Estradiol Affects Epstein–Barr Virus Reactivation-Induced Thyrotropin Receptor Antibody and Immunoglobulin Production in Graves' Disease Patients and Healthy Controls

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Abstract

Epstein–Barr virus (EBV) is a gamma-herpesvirus persisting mainly in human B lymphocytes. EBV reactivation induces host cells to differentiate into plasma cells and is related to autoimmune diseases. Graves' disease, an autoimmune hyperthyroidism, is caused by the thyrotropin receptor antibody (TRAb), which overstimulates thyroid stimulating hormone receptor. The disease occurs predominantly in women, which suggests involvement with estrogen. Graves' disease patients and healthy controls have EBV-infected lymphocytes with TRAb on the surface (TRAb(+)EBV(+) cells) in peripheral blood mononuclear cells (PBMCs). TRAb can be produced by reactivation of EBV *in vitro*, which is an alternative system of antibody production. In this study, we cultured PBMCs from Graves' disease patients and healthy controls with 0, 1, and 100 nM estradiol, corresponding to control, midluteal, and pregnancy levels, respectively, and analyzed the levels of TRAb, total-IgG, and total-IgM during EBV reactivation. We found that 1 nM estradiol increased TRAb levels and 100 nM estradiol slightly lowered them in both patients and controls. In patients, IgM production at 100 nM estradiol was significantly lower than that at 0 nM estradiol ($p=0.028$). Estradiol increased the ratio of IgG production to immunoglobulin G (IgG) and immunoglobulin M (IgM) production (IgG/IgG + IgM), which suggested an increase in class switch recombination in the process of EBV reactivation-induced Ig production. Moreover, TRAb production was stimulated by a midluteal level of estradiol and was suppressed by a pregnancy level of estradiol in controls and patients. These results were consistent with premenstrual worsening and maternity improving of autoimmune diseases, including Graves' disease.

Keywords: estradiol, EBV reactivation, Ig production, Graves' disease

Introduction

MOST AUTOIMMUNE DISEASES predominantly occur in women and are influenced by various factors, including reproductive function, genetics (14,16), and sex hormones (5,6,19,20). Estrogen binds to estrogen receptors (ERs), ER α and ER β , and regulates the menstrual cycle and maternity. Estrogen triggers distinct signaling pathways (6) and can stimulate antibody production from B cells (20). Estradiol is the most potent estrogen. The levels of immunoglobulin G (IgG)

and immunoglobulin M (IgM) in human peripheral blood mononuclear cells (PBMCs) are increased by 0.1–10 nM estradiol and are sustained at concentrations of estradiol higher than 10 nM after 7 days in culture (5).

Nagata *et al.* reported that Graves' disease patients and healthy controls have Epstein–Barr virus (EBV)-infected lymphocytes with thyrotropin receptor antibody (TRAb) on their surface (TRAb(+)EBV(+) cells) in PBMCs, and these PBMCs produce TRAb in response to reactivation of EBV *in vitro* (11,13). Nagata *et al.* also reported that there is a

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positive relationship between serum levels of TRAb and the EBV-early antigen antibody, a marker of EBV reactivation (10). Therefore, EBV reactivation may contribute to auto-antibody production. Graves' disease, an autoimmune hyperthyroidism, is caused by TRAb, which overstimulates thyroid stimulating hormone receptor (8).

We examined the effects of estradiol on antibody production induced by EBV reactivation in PBMCs of healthy controls and Graves' disease patients. We used 0, 1, and 100 nM concentrations of estradiol, corresponding to control, midluteal phase, and pregnancy levels, respectively. We discuss the effects of estradiol levels in the menstrual cycle and pregnancy on EBV reactivation-induced antibody production.

Materials and Methods

Subjects

PBMC samples were obtained from seven Graves' disease patients and eight healthy controls (Table 1). All subjects provided written informed consent, and the study protocol was approved by the Medical Ethics Committee for Human Subject Research (No. 707, 707-1-13) at the Faculty of Medicine, Tottori University, Yonago, Japan. The mean ages ($\pm$ SD) of healthy controls and Graves' disease patients were 29.75 ($\pm$ 10.89) years and 40.00 ($\pm$ 6.22) years, respectively. All subjects were female. At the time of diagnosis, patients exhibited symptoms and had laboratory data that included at least one of the following: (1) signs of thyrotoxicosis such as tachycardia, weight loss, finger tremors, and sweating; (2) diffuse enlargement of the thyroid gland; and (3) exophthalmos and/or specific ophthalmopathy. All patients met the following criteria: (1) elevated serum levels of free T4 and/or free T3; (2) suppression of serum thyrotropin (thyroid stimulating hormone: TSH) ($<0.1 \mu\text{U/mL}$); and (3) positivity for TRAb or thyroid-stimulating antibody. Five of the seven patients were receiving treatments with antithyroid drugs (methylmercaptoimidazole or propylthiouracil). Three

patients were receiving treatments with levothyroxine and No. 11 patient was treated with only levothyroxine. Control subjects were enrolled voluntarily, their thyroid functions were normal, and they had no family history of thyroid disease.

PBMC preparation

Peripheral blood samples were obtained from Graves' disease patients and healthy controls. PBMCs were separated using a Ficoll-Conray density gradient and stored at -80°C until use.

Estradiol

17β -estradiol (Sigma-Aldrich, Saint Louis, MO) was dissolved in ethanol to prepare a 1 mg/mL stock solution and stored at 4°C until use. Estradiol was added to medium and maintained at 0, 1, or 100 nM, corresponding to control, midluteal, or pregnancy levels, respectively. Except for doses of estradiol, every culture was performed with the same medium, and we confirmed that the estradiol concentration of the medium was neglectable.

Sampling protocol

PBMCs were cultured at 37°C for 2 days in RPMI 1640/10% FBS with cyclosporin A ($0.1 \mu\text{g/mL}$) to inhibit T cell effects. PBMCs were transferred to an incubator at 33°C to induce EBV reactivation and were regarded as day 0 samples. On days 0, 5, 10, and 12, half of the culture supernatant was sampled and replaced by fresh medium with estradiol, and the estradiol concentration was maintained at 0, 1, or 100 nM.

Measurement of total immunoglobulin

Immunoglobulin (Ig) concentrations in culture supernatant were measured by ELISA. The total IgG and IgM concentrations were measured using a Human IgG/IgM ELISA Quantitation Set (Bethyl Laboratories, Montgomery, TX).

TABLE 1. SUBJECT PROFILES

	No.	Sex	Age (years)	Allergy	Treatment ^a
Controls	1	F	24	—	—
	2	F	26	—	—
	3	F	22	Allergic conjunctivitis, allergic rhinitis	—
	4	F	27	—	—
	5	F	24	Allergic rhinitis	—
	6	F	22	—	—
	7	F	52	Bronchial asthma	—
	8	F	41	Atopic dermatitis	—
	Mean		29.75		
Patients (Graves' disease)	9	F	48	Atopic dermatitis	MMI 1T, LT4 50 μg
	10	F	39	—	—
	11	F	41	—	LT4 50 μg
	12	F	30	—	PTU 2T
	13	F	36	—	PTU 1T
	14	F	47	—	MMI 1T, LT4 50 μg
	15	F	39	Bronchial asthma, atopic dermatitis	PTU 3T
	Mean		40		

^aTreatment for patients with Graves' disease.

LT4, levothyroxine; MMI, methylmercaptoimidazole; PTU, propylthiouracil.

Determination of TRAb concentration

TRAb concentration in culture fluids was quantified by a radioreceptor assay according to the manufacturer's instructions (DYNOTest TRAb Human; Yamasa Corporation, Choshi, Japan).

Statistical analyses

SPSS Statistics 21 (IBM, Armonk, NY) was used for statistical analyses. A Mann-Whitney *U* test was adopted for comparisons between Graves' disease patients and healthy controls. The Wilcoxon rank sum test was used for the analyses of the effects of estradiol compared with that of baseline (0 nM estradiol).

Results

Effect of estradiol on EBV reactivation-induced Ig production

TRAb production. Compared with TRAb production in the culture at 0 nM estradiol, means of TRAb production were increased with 1 nM estradiol and decreased with 100 nM in healthy controls and patients, although the differences were not statistically significant. There were no significant differences between healthy controls and patients in TRAb production at each estradiol concentration (Fig. 1).

IgG production. The concentration of estradiol did not have a significant effect on IgG production in healthy controls or patients. There were no significant differences in IgG production at each estradiol concentration between healthy controls and patients (Fig. 2).

IgM production. In patients, IgM production at 100 nM estradiol was significantly lower than that at 0 nM estradiol ($p=0.028$). There were no significant differences in IgM

production at each concentration of estradiol between healthy controls and patients (Fig. 3).

Effect of estradiol on the ratio of IgG to IgG and IgM

Figure 4a and b is a pileup of mean IgG and IgM production to overview the ratio of IgG production. Estradiol increased the mean production of IgG in controls (Fig. 4a) and decreased the mean production of IgM in patients (Fig. 4b).

To perform statistical analyses, we calculated the ratio of IgG production to the sum of IgG and IgM production (IgG/IgG + IgM) as for every sample of controls and patients (Fig. 4c, d). In patients, we detected the trend of an increase in the ratio of IgG/IgG + IgM between 0 and 1 nM estradiol ($p=0.063$) and between 0 and 100 nM estradiol ($p=0.128$) (Fig. 4d), but there were no significant differences (Fig. 4d). There were no significant differences in the ratio of IgG/IgG + IgM between healthy controls and patients at each concentration of estradiol (Fig. 5).

Discussion

Similar to previous reports (11–13), we found that EBV reactivation induced production of IgG, IgM, and TRAb in the supernatants of PBMCs from Graves' disease patients and healthy controls (Figs. 1–3). We observed that 1 nM estradiol considerably increased TRAb levels and 100 nM estradiol slightly lowered them in both patients and controls (Fig. 1), although the differences were not statistically significant. TRAb production was stimulated by a midluteal level of estradiol and suppressed by a pregnant level of estradiol, but the difference was not significant probably because there were only a few subjects and large differences in TRAb production between individuals. In a study of menstrual cycle, we often realize the large individual difference probably derived from the number or function of ERs in each subject, as well as the cyclicity of estrogen and progesterone secretion (1–3). The increase in TRAb at midluteal level of estradiol matches

FIG. 1. TRAb production was increased by 1 nM estradiol and decreased by 100 nM estradiol in healthy controls and patients, but differences were not significant. (a) Mean TRAb production in healthy controls. (b) Mean TRAb production in patients. TRAb, thyrotropin receptor antibody.

FIG. 2. The concentration of estradiol did not significantly affect IgG production in healthy controls and patients. (a) Mean IgG production in healthy controls. (b) Mean IgG production in patients. IgG, immunoglobulin G.

premenstrual worsening of autoimmune diseases, including Graves' disease (7,15,18). On the contrary, TRAb suppression by a pregnant level of estradiol is consistent with the improvement of autoimmune diseases, including Graves' disease in maternity (8).

In patients, IgM production at 100 nM estradiol was significantly lower than that at 0 nM estradiol ($p=0.028$), which suggested a relative increase in IgG. In particular, estradiol increased the ratio of IgG/IgG + IgM in both patients and healthy controls. In patients, the IgG/IgG + IgM ratio increased between 0 and 1 nM estradiol ($p=0.063$) and between 0 and 100 nM estradiol ($p=0.128$), but there were no differences in control patients (Fig. 4).

When persistent EBVs are reactivated, host B cells differentiate into plasma cells and produce various isotypes of

Ig, which is the result of class switch recombination (CSR) during Ig production because naive B cells have IgM on their surface. CSR replaces the Ig heavy chain and switches the Ig isotype from IgM to high affinity Ig (9). In EBV reactivation-induced Ig production, expression of activation-induced cytidine deaminase (AID) is induced by EBV-LMP1 (12). Therefore, Ig production involves AID expression and subsequent CSR. An increase in the ratio of IgG/IgG + IgM may result from increased IgG production due to AID induction and CSR. Estrogen binds to the ER and the estrogen-ER complex binds to the *Ig* gene heavy chain locus in B cells, stimulating antibody expression and CSR (4). Moreover, the estrogen-ER complex binds to the AID promoter and increases AID protein production (17). We observed that estradiol increased the ratio of IgG/IgG + IgM,

FIG. 3. IgM production was decreased by 100 nM estradiol. (a) Mean IgM production in healthy controls. (b) Mean IgM production in patients. IgM production at 100 nM estradiol was significantly lower than that at 0 nM estradiol ($p=0.028$). IgM, immunoglobulin M.

FIG. 4. Estradiol increased the ratio of IgG to IgG and IgM (IgG/IgG + IgM) in patients. Pileup production of Ig in healthy controls (a) and patients (b). The ratio of IgG to IgG and IgM (IgG/IgG + IgM) in healthy controls (c) and patients (d). In patients, the IgG/IgG + IgM ratio increased between 0 and 1 nM estradiol ($p=0.063$) and between 0 and 100 nM estradiol ($p=0.128$).

which suggests that estradiol increased CSR in the process of EBV reactivation-induced Ig production.

The ratio of IgG/IgG + IgM in controls was higher than that in patients at all three concentrations of estradiol. However, there were no significant differences between patients and healthy controls at each concentration probably because of a small sample size and large individual differences in controls. EBV reactivation-induced Ig production is influenced by how easily persistent EBV is reactivated. EBV serum antibodies are high in patients with allergies and autoimmune diseases (7,15). Therefore, allergen history is important for selecting control subjects, and examination of how estradiol directly affects EBV reactivation is needed.

In this study, we examined the tonic effect of estradiol on B cell *in vitro* although estradiol may have a tonic and cyclic effect on B cell *in vivo*, for instance, estradiol levels fluctuate during menstrual cycle (1). Therefore, we should investigate further possibility between estradiol and B cell, including the estradiol withdrawal test. Estrogen affects various cells through multiple pathways (2,6). B cells are known to be ER-positive cells, but the receptor number and function may be different individually, similar to mammary glandular cells (1,3). This might be one of the causes of individual difference.

Two of our seven patients (No. 10, 11) were not treated with antithyroid drugs. The levels of TRAb released in culture mediums were lower in No. 11 treated with only

FIG. 5. The effect of estradiol on the ratio of IgG to IgG and IgM (IgG/IgG + IgM) in healthy controls and patients. There were no significant differences between healthy controls and patients for each concentration of estradiol.

levothyroxine than the means of patients in every estradiol concentration, but in No. 10 without treatment, they were higher.

Antithyroid drugs are reported to have immunomodulatory effects and that is because the patients receiving antithyroid drug show lowered serum TRAb levels (clinical observation). These are also future tasks.

In conclusion, we examined the effect of estradiol on EBV reactivation-induced Ig production, an alternative system of antibody production. Because of a small sample size and individual difference, including the numbers and function of ER, we did not observe sufficiently significant differences. However, we detected a significant decrease in IgM production at 100 nM estradiol in patients and showed the trend that estradiol increased the ratio of IgG/IgG + IgM, which suggests that estradiol increased CSR. Furthermore, 1 nM estradiol (the midluteal level) may have the effect to increase TRAb production and 100 nM estradiol (the pregnancy level) may suppress TRAb production. These results are consistent with clinical observations of premenstrual worsening and maternity improving of autoimmune disease, including Graves' disease (7,15,18).

Acknowledgments

We are grateful to the laboratory staff of Molecular Pathology Division, Tottori University. This work was supported by JSPS KAKENHI Grant No. 17K08694 (K.N.).

Author Disclosure Statement

No competing financial interests exist.

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